

**Immunohistochemistry**

Policy Number: AHS –P2018 – Immunohistochemistry	Prior Policy Name and Number, as applicable:
Effective Date: 05/01/2024	

[POLICY DESCRIPTION](#) | [INDICATIONS AND/OR LIMITATIONS OF COVERAGE](#) | [TABLE OF TERMINOLOGY](#) | [SCIENTIFIC BACKGROUND](#) | [GUIDELINES AND RECOMMENDATIONS](#) | [APPLICABLE STATE AND FEDERAL REGULATIONS](#) | [APPLICABLE CPT/HCPCS PROCEDURE CODES](#) | [EVIDENCE-BASED SCIENTIFIC REFERENCES](#) | [REVISION HISTORY](#)

**I. Policy Description**

Immunohistochemistry (IHC) is a very sensitive and specific staining technique that uses anatomical, biochemical, and immunological methods to identify cells, tissues, and organisms by the interaction of target antigens with highly specific monoclonal antibodies and visualization through the use of a biochemical tag or label (Fitzgibbons et al., 2014).

**II. Indications and/or Limitations of Coverage**

Application of coverage criteria is dependent upon an individual’s benefit coverage at the time of the request.

- 1) Code 88342 should be used for the first single antibody procedure and is reimbursed at one unit per specimen, up to four specimens, per date of service.
- 2) Code 88341 should be used for each additional single antibody per specimen and is reimbursed up to a maximum of 13 units per date of service.
- 3) Code 88344 should be used for each multiplex antibody per specimen, up to six specimens, per date of service.

**III. Table of Terminology**

Term	Definition
AFP	Alpha-fetoprotein
ARID1A	AT-rich interactive domain-containing protein 1A
ASCO	The American Society of Clinical Oncology
<i>Bcl2</i>	<i>BCL2 apoptosis regulator</i>
b-HCG	Beta human chorionic gonadotropin
<i>BRCA1</i>	Breast cancer type 1 susceptibility protein gene
BAP1	<i>BRCA1</i> associated protein 1
CAIX	Carbonic anhydrase IX
CAP	College of American Pathologists
CD1a	Cluster of differentiation 1a

CD5	Cluster of differentiation 5
CD10	Cluster of differentiation 10
CD21	Cluster of differentiation 21
CD30	Cluster of differentiation 30
CD31	Cluster of differentiation 31
CD34	Cluster of differentiation 34
CD35	Cluster of differentiation 35
CD43	Cluster of differentiation 43
CD56	Cluster of differentiation 56
CD99	Cluster of differentiation 99
CD117	Cluster of differentiation 117
<i>CDH17</i>	<i>Cadherin-17</i>
<i>CDK4</i>	<i>Cyclin-dependent kinase 4</i>
CDX2	Caudal-type homeobox 2
CEA	Carcinoembryonic antigen
CK	Creatine kinase
CK17	Cytokeratin 17
CK20	Cytokeratin 20
CK5/6	Cytokeratin 5/6
CK903	Cytokeratin 903
CLIA'88	Clinical Laboratory Improvement Amendments of 1988
CMS	Centres for Medicare and Medicaid
CRC	Colorectal cancer
D2-40	Anti-Podoplanin
DNA	Deoxyribonucleic acid
DOG1	Delay of germination 1
ERG	ETS-related gene
ESMO	The European Society of Medical Oncology
FDA	Food and Drug Administration
FISH	Fluorescence in situ hybridization
Fli-1	Friend leukemia integration 1
FOXL2	Forkhead box protein L2
GATA3	GATA binding protein 3
GCDFP15	Gross cystic disease fluid protein 15
GI	Gastrointestinal tract
HepPar-1	General hepatocyte paraffin 1
<i>HER2</i>	<i>Human epidermal growth factor receptor 2</i>
HMB-45	Human melanoma black-45
HNF-1b	Hepatocyte nuclear factor 1 beta
HPV	Human papillomavirus
IHC	Immunohistochemistry
IMP3	U3 small nucleolar ribonucleoprotein protein IMP3
INI1	Integrase interactor 1

ISH	In situ hybridization
KIM-1	Kidney injury molecule-1
LDTs	Laboratory developed tests
Maspin	Mammary serine protease inhibitor
MCPyV	Merkel cell polyomavirus
MDM2	Mouse double minute 2 homolog
MIB-1	MIB E3 ubiquitin protein ligase 1
mIHC	Multiplex immunohistochemistry
MiTF	Microphthalmia-associated transcription factor
MLH1	MutL homolog 1
MMR	Mismatch repair protein
MPO	Myeloperoxidase
MSA	Mammary serum antigen
MSH2	Mismatch repair protein Msh2
MSI	Microsatellite instability
MUC4	Mucin 4
MUC5AC	Mucin 5AC
MyoD1	Myoblast determination protein 1
<i>NANOG</i>	<i>Nanog Homeobox</i>
napsin A	Novel aspartic proteinase of the pepsin family A
NCCN	The National Cancer Coalition Network
NKX2.2	Homeobox protein
NKX3.1	Homeobox protein
NY-ESO-1	New York esophageal squamous cell carcinoma 1
OCT4	Octamer-binding transcription factor 4
p16	Cyclin-dependent kinase inhibitor 2A
p40	Protein subunit
P504S	Cytoplasmic protein
p63	Tumor protein p63
pan-Trk	Pan-tropomyosin-related-kinase
PAX2	Paired box 2
PAX8	Paired box 8
PDX1	Insulin promoter factor 1
PNET	Primitive neuro-ectodermal tumor
PSA	Prostate-specific antigen
PSAP	Phosphoserine aminotransferase
PTEN	Phosphatase and tensin homolog
pVHL	Von hippel–lindau tumor suppressor
RB	Retinoblastoma protein
RCC	Renal cell carcinoma
RCCma	Renal cell carcinoma marker
S100P	S100 calcium-binding protein p
SALL4	Sal-like protein 4

SATB2	Special AT-rich sequence-binding protein 2
SF-1	Steroidogenic factor 1
SOX10	SRY-box transcription factor 10
TFE3	Transcription factor E3
TLE1	Transducin-like enhancer protein 1
TTF1	Transcription termination factor, RNA polymerase I
UPII	Uroplakin II
WT1	Wilms tumor protein

#### IV. Scientific Background

Immunohistochemistry (IHC) is used to identify certain components of tissues or cells (aka immunocytochemistry) via use of specific antibodies that can be visualized through a staining technique. The premise behind IHC is that distinct tissues and cells contain a unique set of antigens that allows them to be identified and differentiated. The selection of antibodies used for the evaluation of a specimen varies by the source of the specimen, the question to be answered, and the pathologist performing the test.

Importantly, an entirely sensitive and specific IHC marker rarely exists, and therefore, determinations are typically based on a pattern of positive and negative stains for a panel of several antibodies. The four most common IHC staining patterns include nuclear staining, cytoplasmic staining, membrane staining, and extracellular staining (Tuffaha et al., 2018). A single IHC marker approach (other than for pathogens such as cytomegalovirus or BK virus) is strongly discouraged since aberrant expression of a highly specific IHC marker can rarely occur. However, aberrant expression of the entire panel of highly specific IHC markers is nearly statistically impossible (Lin & Chen, 2014).

Multiplex immunohistochemistry (mIHC) is a particular IHC technique that allows multiple targets in a single tissue to be detected simultaneously; this approach is able to characterize “the tumor microenvironment including vascular architecture and hypoxia, cellular proliferation, cell death as well as drug distribution” (Kalra & Baker, 2017). Hence, mIHC can assist in the development of parameter tumor maps. Other researchers have utilized mIHC for its novel ability to provide quantitative data on different types of tumor-infiltrating immune cells within a single tissue; this may improve cancer patient immunotherapy stratification (Hofman et al., 2019).

#### *Clinical Utility and Validity*

Immunohistochemistry can be used for a variety of purposes including: differentiation of benign from malignant tissue, differentiation among several types of cancer, selection of therapy, identification of the origin of a metastatic cancer, and identification of infectious organisms (Shah et al., 2012). IHC has many uses in the realm of tumor identification, and it has even been clinically used to pinpoint various breast cancer-specific markers, such as progesterone and estrogen receptors, gross cystic duct fluid protein, and mammaglobin (Hainsworth & Greco, 2022). Further, overexpression of the *HER2* oncogene, a predictive breast cancer biomarker, is often identified via IHC (Yamauchi & Bleiweiss, 2023). In regards to tumor identification, a specific type of IHC, known as pan-Trk IHC, has been shown to positively identify inflammatory

myofibroblastic tumors with a nuclear and cytoplasmic staining pattern that may assist in targeted therapy (Yamamoto et al., 2019).

Antibodies for use in IHC are available as single antibody reagents or in mixtures of a combination of antibodies. More than 200 diagnostic antibodies are generally available in a large clinical IHC laboratory, and hundreds of antibodies are usually available in research laboratories. The list of new antibodies is growing rapidly with the discovery of new biomarkers by molecular methodologies (Lizotte et al., 2016). Several studies have shown that a relatively low number of antibodies are capable of accurately diagnosing specific cancers and identifying the primary source of a metastasis (Le Stang et al., 2019; Lizotte et al., 2016; Prok & Prayson, 2006).

Common markers to identify tumor origin (Lin & Chen, 2014):

<b>Primary Site</b>	<b>Markers</b>
<b>Lung adenocarcinoma</b>	TTF1, napsin A
<b>Breast carcinoma</b>	GATA3, ER, GCDFP15
<b>Urothelial carcinoma</b>	GATA3, UPII, S100P, CK903, p63
<b>Squamous cell carcinoma</b>	p40, CK5/6
<b>RCC, clear cell type</b>	PAX8, RCCma, pVHL, KIM-1
<b>Papillary RCC</b>	P504S, RCCma, pVHL, PAX8, KIM-1
<b>Translocational RCC</b>	TFE3
<b>Hepatocellular carcinoma</b>	Arginase-1, glypican-3, HepPar-1
<b>Adrenal cortical neoplasm</b>	Mart-1, inhibin-a, calretinin, SF-1
<b>Melanoma</b>	S100, Mart-1, HMB-45, MiTF, SOX10
<b>Merkel cell carcinoma</b>	CK20 (perinuclear dot staining), MCPyV
<b>Mesothelial origin</b>	Calretinin, WT1, D2-40, CK5/6, mesothelin
<b>Neuroendocrine origin</b>	Chromogranin, synaptophysin, CD56
<b>Upper GI tract</b>	CDH17, CDX2, CK20
<b>Lower GI tract</b>	CDH17, SATB2, CDX2, CK20
<b>Intrahepatic cholangiocarcinoma</b>	pVHL, CAIX
<b>Pancreas, acinar cell carcinoma</b>	Glypican-3, antitrypsin
<b>Pancreas, ductal adenocarcinoma</b>	MUC5AC, CK17, Maspin, S100P, IMP3
<b>Pancreas, neuroendocrine tumor</b>	PR, PAX8, PDX1, CDH17, islet-1
<b>Pancreas, solid pseudopapillary tumor</b>	Nuclear b-catenin, loss of Ecadherin, PR, CD10, vimentin
<b>Prostate, adenocarcinoma</b>	PSA, NKX3.1, PSAP, ERG

<b>Ovarian serous carcinoma</b>	PAX8, ER, WT1
<b>Ovarian clear cell carcinoma</b>	pVHL, HNF-1b, KIM-1, PAX8
<b>Endometrial stromal sarcoma</b>	CD10, ER
<b>Endometrial adenocarcinoma</b>	PAX8/PAX2, ER, vimentin
<b>Endocervical adenocarcinoma</b>	PAX8, p16, CEA, HPV in situ hybridization, loss of PAX2
<b>Thyroid follicular cell origin</b>	TTF1, PAX8, thyroglobulin
<b>Thyroid medullary carcinoma</b>	Calcitonin, TTF1, CEA
<b>Hyalinizing trabecular adenoma of the thyroid</b>	MIB-1 (unique membranous staining pattern)
<b>Salivary duct carcinoma</b>	GATA3, AR, GCDFP-15, HER2/neu
<b>Thymic origin</b>	PAX8, p63, CD5
<b>Seminoma</b>	SALL4, OCT4, CD117, D2-40
<b>Yolk sac tumor</b>	SALL4, glypican-3, AFP
<b>Embryonal carcinoma</b>	SALL4, OCT4, NANOG, CD30
<b>Choriocarcinoma</b>	b-HCG, CD10, SALL4
<b>Sex cord–stromal tumors</b>	SF-1, inhibin-a, calretinin, FOXL2
<b>Vascular tumor</b>	ERG, CD31, CD34, Fli-1
<b>Synovial sarcoma</b>	TLE1, cytokeratin
<b>Chordoma</b>	Cytokeratin, S100
<b>Desmoplastic small round cell tumor</b>	Cytokeratin, CD99, desmin, WT1 (N-terminus)
<b>Alveolar soft part sarcoma</b>	TFE3
<b>Rhabdomyosarcoma</b>	Myogenin, desmin, MyoD1
<b>Smooth muscle tumor</b>	SMA, MSA, desmin, calponin
<b>Ewing sarcoma/PNET</b>	NKX2.2, CD99, Fli-1
<b>Myxoid and round cell liposarcoma</b>	NY-ESO-1
<b>Low-grade fibromyxoid sarcoma</b>	MUC4
<b>Epithelioid sarcoma</b>	Loss of INI1, CD34, CK
<b>Atypical lipomatous tumor</b>	MDM2 (MDM2 by FISH is a more sensitive and specific test), CDK4
<b>Histiocytosis X</b>	CD1a, S100

<b>Angiomyolipoma</b>	HMB-45, SMA
<b>Gastrointestinal stromal tumor</b>	CD117, DOG1
<b>Solitary fibrous tumor</b>	CD34, Bcl2, CD99
<b>Myoepithelial carcinoma</b>	Cytokeratin and myoepithelial markers; may lose INI1
<b>Myeloid sarcoma</b>	CD43, CD34, MPO
<b>Follicular dendritic cell tumor</b>	CD21, CD35
<b>Mast cell tumor</b>	CD117, tryptase

## V. Guidelines and Recommendations

Guidelines are lacking regarding the selection and number of antibodies that should be used for most immunohistochemistry evaluations. However, IHC is broadly used for conditions such as cancers, which are mentioned across many different societies. The below section is not a comprehensive list of guidance for immunohistochemistry.

### **College of American Pathologists (CAP)**

The College of American Pathologists has published several reviews in Archives of Pathology & Laboratory Medicine that detail the quality control measures for IHC; further, CAP has also published more than 100 small IHC panels to address the frequently asked questions in diagnosis and differential diagnosis of specific entities. These diagnostic panels are based on literature, IHC data, and personal experience. A single IHC marker approach (other than for pathogens such as cytomegalovirus or BK virus) is strongly discouraged since aberrant expression of a highly specific IHC marker can rarely occur. However, aberrant expression of the entire panel of highly specific IHC markers is nearly statistically impossible (Lin & Chen, 2014; Lin & Liu, 2014).

### **The American Society of Clinical Oncology (ASCO) and the College of American Pathologists (CAP)**

The American Society of Clinical Oncology and the College of American Pathologists currently recommend that “all newly diagnosed patients with breast cancer must have a HER2 test performed” (Wolff et al., 2013). Also, for those who develop metastatic disease, a HER2 test must be done on tissue from the metastatic site, if available. In less common HER2 breast cancer patterns, as observed in approximately 5% of cases by dual-probe in situ hybridization (ISH) assays, new recommendations have been made to make a final determination of positive or negative HER2 tissue. This new “diagnostic approach includes more rigorous interpretation criteria for ISH and requires concomitant IHC review for dual-probe ISH groups... to arrive at the most accurate HER2 status designation (positive or negative) based on combined interpretation of the ISH and IHC assays;” further, “The Expert Panel recommends that laboratories using single-probe ISH assays include concomitant IHC review as part of the interpretation of all single-probe ISH assay results” (Wolff et al., 2018).

The 2018 update included the following changes from the prior 2013 update, particularly focusing on infrequent HER2 test results that were of “uncertain biologic or clinical significance”:

- “Revision of the definition of IHC 2+ (equivocal) to the original FDA-approved criteria.
- Repeat HER2 testing on a surgical specimen if the initially tested core biopsy is negative is no longer stated as mandatory. A new HER2 test *may* (no longer *should*) be ordered on the excision specimen on the basis of some criteria (such as tumor grade 3).
- A more rigorous interpretation criteria of the less common patterns that can be seen in about 5% of all cases when HER2 status in breast cancer is evaluated using a dual-probe ISH testing. These cases, described as ISH groups 2 to 4, should now be assessed using a diagnostic approach that includes a concomitant review of the IHC test, which will help the pathologist make a final determination of the tumor specimen as HER2 positive or negative.

The Expert Panel also preferentially recommends the use of dual-probe instead of single-probe ISH assays, but it recognizes that several single-probe ISH assays have regulatory approval in many parts of the world” (Wolff et al., 2018). The 2018 recommendations were affirmed in 2023 (Wolff et al., 2023).

### **The National Cancer Coalition Network**

The NCCN has made numerous recommendations for use of IHC to diagnose and manage various types of cancer. Cancers with clinically useful IHC applications include breast, cervical, various leukemias, and colorectal cancer.

The NCCN states that the determination of estrogen receptor, progesterone receptor, and HER2 status for breast cancer is recommended and may be determined by IHC (NCCN, 2023a). Specifically, the guidelines state that “the NCCN Panel endorses the CAP protocol for pathology reporting and endorses the ASCO CAP recommendations for quality control performance of HER2 testing and interpretation of IHC and ISH results.” They also specifically endorse the ASCO/CAP HER2 testing guideline “Principles of HER2 testing,” and state “HR testing (ER and PR) by IHC should be performed on any new primary or newly metastatic breast cancer using methodology outlined in the latest ASCO/CAP HR testing guideline.” Additionally, “PR testing by IHC on invasive cancers can aid in the prognostic classification of cancers and serve as a control for possible false negative ER results. Patients with ER-negative, PR-positive cancers may be considered for endocrine therapies, but the data on this group are noted to be limited” (NCCN, 2023a).

Further, the NCCN recommendations concerning genetic testing for colorectal cancer state, “The panel recommends that for patients or families where colorectal or endometrial tumor is available, one of three options should be considered for workup: 1) tumor testing with IHC or MSI; 2) comprehensive NGS panel (that includes, at minimum, the four MMR genes and *EPCAM*, *BRAF*, MSI, and other known familial cancer genes); or 3) germline multi-gene testing that includes the four MMR genes and *EPCAM*. The panel recommends tumor testing with IHC and/or MSI be used as the primary approach for pathology-lab-based universal screening” (NCCN, 2023b). More recently, the NCCN has made additional recommendations to individuals diagnosed with any type of hereditary colorectal cancer (CRC) syndrome; these



recommendations state that “all individuals newly diagnosed with CRC have either MSI or immunohistochemistry (IHC) testing for absence of 1 of the 4 DNA MMR proteins” (NCCN, 2023b).

### The European Society of Medical Oncology (ESMO)

The ESMO recommends that for cancers of an unknown primary site, “histology and IHC on good quality tissue specimens are required [III, A]” (Krämer et al., 2023). Particularly in the context for gastrointestinal carcinomas, ESMO states “Immunohistochemical loss of *BRCAl*-associated protein 1 (BAP1) or AT-rich interactive domain-containing protein 1A (ARID1A) can support the diagnosis but the final decision can only be made in conjunction with clinical and radiological findings.” Other mentions of IHC in their updated 2023 guidelines did not result in any other updated recommendations (Krämer et al., 2023).

## VI. Applicable State and Federal Regulations

### Food and Drug Administration (FDA)

Many labs have developed specific tests that they must validate and perform in house. These laboratory-developed tests (LDTs) are regulated by the Centers for Medicare and Medicaid (CMS) as high-complexity tests under the Clinical Laboratory Improvement Amendments of 1988 (CLIA '88). LDTs are not approved or cleared by the U. S. Food and Drug Administration; however, FDA clearance or approval is not currently required for clinical use.

Recently, four clinical IHC biomarker assays (PTEN, RB, MLH1, and MSH2) have been validated for use as biomarkers in a nationwide clinical trial; these assays were then approved by the FDA as laboratory-developed tests to assist in the treatment selection of patients in clinical trials (Khoury et al., 2018). This shows that IHC assays are currently being utilized with molecular tests to assist in therapeutic decisions.

## VII. Applicable CPT/HCPCS Procedure Codes

Procedure codes appearing in medical policy documents are only included as a general reference. This list may not be all inclusive and is subject to updates. In addition, codes listed are not a guarantee of payment.

CPT	Code Description
88341	Immunohistochemistry or immunocytochemistry, per specimen; each additional single antibody stain procedure
88342	Immunohistochemistry or immunocytochemistry, per spec; initial single antibody stain
88344	Immunohistochemistry or immunocytochemistry, per specimen; each multiplex antibody stain procedure

Current Procedural Terminology<sup>®</sup> American Medical Association. All Rights reserved.

## VIII. Evidence-based Scientific References

- Fitzgibbons, P. L., Bradley, L. A., Fatheree, L. A., Alsabeh, R., Fulton, R. S., Goldsmith, J. D., Haas, T. S., Karabakhtsian, R. G., Loykasek, P. A., Marolt, M. J., Shen, S. S., Smith, A. T., & Swanson, P. E. (2014). Principles of analytic validation of immunohistochemical assays: Guideline from the College of American Pathologists Pathology and Laboratory Quality Center. *Arch Pathol Lab Med*, 138(11), 1432-1443. <https://doi.org/10.5858/arpa.2013-0610-CP>
- Hainsworth, J., & Greco, F. (2022, January 20). *Overview of the classification and management of cancers of unknown primary site*. <https://www.uptodate.com/contents/overview-of-the-classification-and-management-of-cancers-of-unknown-primary-site>
- Hofman, P., Badoual, C., Henderson, F., Berland, L., Hamila, M., Long-Mira, E., Lassalle, S., Roussel, H., Hofman, V., Tartour, E., & Ilie, M. (2019). Multiplexed Immunohistochemistry for Molecular and Immune Profiling in Lung Cancer-Just About Ready for Prime-Time? *Cancers (Basel)*, 11(3). <https://doi.org/10.3390/cancers11030283>
- Kalra, J., & Baker, J. (2017). Multiplex Immunohistochemistry for Mapping the Tumor Microenvironment. *Methods Mol Biol*, 1554, 237-251. [https://doi.org/10.1007/978-1-4939-6759-9\\_17](https://doi.org/10.1007/978-1-4939-6759-9_17)
- Khoury, J. D., Wang, W. L., Prieto, V. G., Medeiros, L. J., Kalhor, N., Hameed, M., Broaddus, R., & Hamilton, S. R. (2018). Validation of Immunohistochemical Assays for Integral Biomarkers in the NCI-MATCH EAY131 Clinical Trial. *Clin Cancer Res*, 24(3), 521-531. <https://doi.org/10.1158/1078-0432.Ccr-17-1597>
- Krämer, A., Bochtler, T., Pauli, C., Baciarello, G., Delorme, S., Hemminki, K., Mileschkin, L., Moch, H., Oien, K., Olivier, T., Patrikidou, A., Wasan, H., Zarkavelis, G., Pentheroudakis, G., & Fizazi, K. (2023). Cancer of unknown primary: ESMO Clinical Practice Guideline for diagnosis, treatment and follow-up. *Ann Oncol*, 34(3), 228-246. <https://doi.org/10.1016/j.annonc.2022.11.013>
- Le Stang, N., Burke, L., Blaizot, G., Gibbs, A. R., Lebailly, P., Clin, B., Girard, N., & Galateau-Salle, F. (2019). Differential Diagnosis of Epithelioid Malignant Mesothelioma With Lung and Breast Pleural Metastasis: A Systematic Review Compared With a Standardized Panel of Antibodies-A New Proposal That May Influence Pathologic Practice. *Arch Pathol Lab Med*. <https://doi.org/10.5858/arpa.2018-0457-OA>
- Lin, F., & Chen, Z. (2014). Standardization of diagnostic immunohistochemistry: literature review and geisinger experience. *Arch Pathol Lab Med*, 138(12), 1564-1577. <https://doi.org/10.5858/arpa.2014-0074-RA>
- Lin, F., & Liu, H. (2014). Immunohistochemistry in undifferentiated neoplasm/tumor of uncertain origin. *Arch Pathol Lab Med*, 138(12), 1583-1610. <https://doi.org/10.5858/arpa.2014-0061-RA>
- Lizotte, P. H., Ivanova, E. V., Awad, M. M., Jones, R. E., Keogh, L., Liu, H., Dries, R., Almonte, C., Herter-Sprie, G. S., Santos, A., Feeney, N. B., Pawletz, C. P., Kulkarni, M. M., Bass, A. J., Rustgi, A. K., Yuan, G. C., Kufe, D. W., Janne, P. A., Hammerman, P. S., . . . Wong, K. K. (2016). Multiparametric profiling of non-small-cell lung cancers reveals distinct immunophenotypes. *JCI Insight*, 1(14), e89014. <https://doi.org/10.1172/jci.insight.89014>
- NCCN. (2023a, March 23). *NCCN Clinical Practice Guidelines in Oncology: Breast Cancer Version 4.2023*. National Comprehensive Cancer Network. [https://www.nccn.org/professionals/physician\\_gls/pdf/breast.pdf](https://www.nccn.org/professionals/physician_gls/pdf/breast.pdf)

- NCCN. (2023b, May 30). *NCCN Clinical Practice Guidelines in Oncology: Genetic/Familial High-Risk Assessment: Colorectal Version 1.2023*.  
[https://www.nccn.org/professionals/physician\\_gls/pdf/genetics\\_colon.pdf](https://www.nccn.org/professionals/physician_gls/pdf/genetics_colon.pdf)
- Prok, A. L., & Prayson, R. A. (2006). Thyroid transcription factor–1 staining is useful in identifying brain metastases of pulmonary origin. *Annals of Diagnostic Pathology*, *10*(2), 67-71. <https://doi.org/10.1016/j.anndiagpath.2005.07.013>
- Shah, A. A., Frierson, H. F., & Cathro, H. P. (2012). Analysis of Immunohistochemical Stain Usage in Different Pathology Practice Settings.  
<https://doi.org/10.1309/AJCPAGVTCKDXKK0X>
- Tuffaha, M. S. A., Guski, H., & Kristiansen, G. (2018). Immunohistochemistry in Tumor Diagnostics. In M. S. A. Tuffaha, H. Guski, & G. Kristiansen (Eds.), *Immunohistochemistry in Tumor Diagnostics* (pp. 1-9). Springer International Publishing.  
[https://doi.org/10.1007/978-3-319-53577-7\\_1](https://doi.org/10.1007/978-3-319-53577-7_1)
- Wolff, A. C., Hammond, M. E., Hicks, D. G., Dowsett, M., McShane, L. M., Allison, K. H., Allred, D. C., Bartlett, J. M., Bilous, M., Fitzgibbons, P., Hanna, W., Jenkins, R. B., Mangu, P. B., Paik, S., Perez, E. A., Press, M. F., Spears, P. A., Vance, G. H., Viale, G., & Hayes, D. F. (2013). Recommendations for human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists clinical practice guideline update. *J Clin Oncol*, *31*(31), 3997-4013.  
<https://doi.org/10.1200/jco.2013.50.9984>
- Wolff, A. C., Hammond, M. E. H., Allison, K. H., Harvey, B. E., Mangu, P. B., Bartlett, J. M. S., Bilous, M., Ellis, I. O., Fitzgibbons, P., Hanna, W., Jenkins, R. B., Press, M. F., Spears, P. A., Vance, G. H., Viale, G., McShane, L. M., & Dowsett, M. (2018). Human Epidermal Growth Factor Receptor 2 Testing in Breast Cancer: American Society of Clinical Oncology/College of American Pathologists Clinical Practice Guideline Focused Update. *J Clin Oncol*, *36*(20), 2105-2122. <https://doi.org/10.1200/jco.2018.77.8738>
- Wolff, A. C., Somerfield, M. R., Dowsett, M., Hammond, M. E. H., Hayes, D. F., McShane, L. M., Saphner, T. J., Spears, P. A., & Allison, K. H. (2023). Human Epidermal Growth Factor Receptor 2 Testing in Breast Cancer: ASCO–College of American Pathologists Guideline Update. *Journal of Clinical Oncology*, *41*(22), 3867-3872.  
<https://doi.org/10.1200/JCO.22.02864>
- Yamamoto, H., Nozaki, Y., Kohashi, K., Kinoshita, I., & Oda, Y. (2019). Diagnostic utility of pan-Trk immunohistochemistry for inflammatory myofibroblastic tumors. *Histopathology*.  
<https://doi.org/10.1111/his.14010>
- Yamauchi, H., & Bleiweiss, I. (2023, August 25). *HER2 and predicting response to therapy in breast cancer*. <https://www.uptodate.com/contents/her2-and-predicting-response-to-therapy-in-breast-cancer>

## IX. Revision History

Revision Date	Summary of Changes
01/01/2022	Initial Effective Date
05/23/2022	Updated background, guidelines, and evidence-based scientific references. Literature review did not necessitate any modifications to the coverage criteria.

04/04/2023	Updated the background, guidelines and recommendations, and evidence-based scientific references. Literature review did not necessitate any modifications to coverage criteria. Committee approved 04/04/2023
02/12/2024	Reviewed and Updated: Updated the background, guidelines and recommendations, and evidence-based scientific references. Literature review did not necessitate any modifications to coverage criteria. Committee approved: 02/12/2024